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(54) Title: IONIC MOLECULAR CONJUGATES OF BIODEGRADABLE POLYESTERS AND BIOACTIVE POLYPEPTIDES			
(57) Abstract			
<p>Disclosed is a sustained release pharmaceutical composition. The composition includes a polyester containing a free COOH group ionically conjugated with a bioactive polypeptide comprising at least one effective ionogenic amine, wherein at least 50 % by weight, of the polypeptide present in the composition is ionically conjugated to the polyester.</p>			

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IONIC MOLECULAR CONJUGATES
OF BIODEGRADABLE POLYESTERS AND BIOACTIVE POLYPEPTIDES

Background of the Invention

5 This invention pertains to sustained release of bioactive polypeptides.

 Many drug delivery systems have been developed, tested and utilized for the controlled in vivo release of pharmaceutical compositions. For example, 10 polyesters such as poly (DL-lactic acid), poly (glycolic acid), poly(ϵ - caprolactone) and various other copolymers have been used to release biologically active molecules such as progesterone; these have been in the form of microcapsules, films, or rods (Pitt CG, Marks, TA, and 15 Schindler, A. 1980). Upon implantation of the polymer/therapeutic agent composition, for example subcutaneously or intramuscularly, the therapeutic agent is released over a specific period of time. Such biocompatible biodegradable polymeric systems are 20 designed to permit the entrapped therapeutic agent to diffuse from the polymer matrix. Upon release of the therapeutic agent, the polymer is degraded in vivo, obviating surgical removal of the implant. Although the factors that contribute to polymer degradation are not 25 well understood, it is believed that such degradation for polyesters may be regulated by the accessibility of ester linkages to non-enzymatic autocatalytic hydrolysis of the polymeric components.

 Several EPO publications and US Patents have 30 addressed issues of polymer matrix design and its role in regulating the rate and extent of release of therapeutic agents in vivo.

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For example, Deluca (EPO Publication 0 467 389 A2/Univ of Kentucky) describes a physical interaction between a hydrophobic biodegradable polymer and a protein or polypeptide. The composition formed was a mixture of
5 a therapeutic agent and a hydrophobic polymer that sustained its diffusional release from the matrix after introduction into a subject.

Hutchinson (US Patent 4, 767,628/ICI) controlled the release of a therapeutic agent by uniform dispersion
10 in a polymeric device. It is stated that this formulation provides for controlled continuous release by the overlap of two phases: first, a diffusion-dependent leaching of the drug from the surface of the formulation; and second, releasing by aqueous channels induced by
15 degradation of the polymer.

Summary of the Invention

In general, the invention features a sustained release pharmaceutical formulation composed of a polyester containing one or more free COOH groups
20 ionically conjugated with a biologically active polypeptide composed of at least one effective, ionogenic amine wherein at least 50%, by weight, of the polypeptide present in the composition is ionically conjugated to the polyester.

25 In preferred embodiments, the polyester is modified to increase the carboxyl to hydroxyl end group ratio from more than one and approaches infinity, i.e., all of the hydroxyl groups can be substituted with carboxyls. Examples of suitable polyesters are those
30 originating from compounds such as L-lactic acid, D-lactic acid, DL-lactic acid, ϵ -caprolactone, p-dioxanone, ϵ -caproic acid, substituted and unsubstituted trimethylene carbonate, 1,5-dioxepan-2-one, 1,4-dioxepan-2-one, glycolide, glycolic acid, L-lactide, D-lactide,

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DL-lactide, meso-lactide, alkylene oxalate, cycloalkylene oxalate, alkylene succinate, (β -hydroxybutyrate), and optically active isomers, racemates or copolymers of any of the above. Other heterochain polymers related to
5 traditional polyesters may also be used (e.g., polyorthoesters, polyorthocarbonates, and polyacetals).

Preferably, the polyester is made polycarboxylic by reaction with malic acid or citric acid.

In preferred embodiments the polyester is
10 partially acid-tipped with glutaric anhydride. In yet other preferred embodiments the polyester is fully acid-tipped with glutaric anhydride. Preferably, the polyester has an average degree of polymerization between 10 and 300 and more preferably between 20 to 50.

15 The ionic molecular conjugates of the invention are preferably made from polycarboxylic acid-tipped polyesters conjugated with monobasic and polybasic bioactive polypeptides having at least one effective ionogenic amine group. Alternatively, any polyester may
20 be used to form an ionic molecular conjugate of the invention provided it is pretreated with a suitable base, e.g., NaOH. Moreover, any acid-stable peptide can be used, e.g., growth hormone releasing peptide (GHRP), luteinizing hormone-releasing hormone (LHRH),
25 somatostatin, bombesin, gastrin releasing peptide (GRP), calcitonin, bradykinin, galanin, melanocyte stimulating hormone (MSH), growth hormone releasing factor (GRF), amylin, tachykinins, secretin, parathyroid hormone (PTH), enkaphelin, endothelin, calcitonin gene releasing peptide
30 (CGRP), neuromedins, parathyroid hormone related protein (PTHrP), glucagon, neurotensin, adrenocorticotrophic hormone (ACTH), peptide YY (PYY), glucagon releasing peptide (GLP), vasoactive intestinal peptide (VIP), pituitary adenylate cyclase activating peptide (PACAP),
35 motilin, substance P, neuropeptide Y (NPY), TSH, and

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analogues and fragments thereof. Such ionic molecular conjugates are capable of releasing their bioactive components in vivo at predetermined rates determined by the chemical structure, molecular weight, and pKa of both components of these conjugates. A mechanism for the release of the drug entails the transformation of the insoluble conjugate form to water-soluble components, in part, through the hydrolysis of the hydrophobic polyester. Thus, release of the bioactive polypeptide increases, independently, with (a) the decrease in the pKa differential between the bioactive polypeptide and polyester, (b) the chemical reactivity of the polyester chain that is reflected in the carbonyl nucleophilicity, (c) the decrease in polyester density, as it relates to the glass transition temperature and minimized crystallizability, and (d) the increase in matrix hydrophilicity.

In preferred embodiments the polypeptide comprises 1 to 50 percent by weight of the total weight of the ionic molecular conjugate, and preferably greater than 85%, more preferably 95%, and even more preferably 99%, of the polypeptide present in the composition is ionically conjugated to the polyester; the polyester component of the ionic molecular conjugate has a viscosity of about 0.05 to about 0.7 dl/gm in chloroform; and the polyester has an average molecular weight of about 1200 - 40,000.

The polymeric ionic molecular conjugates of the invention can easily be made into injectable microspheres or microparticles, and implantable films or rods, without the need to utilize processing that entails multiphase emulsions or non-aqueous two-phase systems. Preferably, the microparticles are manufactured by (a) dissolving the composition in an aprotic, water miscible organic solvent; (b) mixing the organic solvent in water; and (c)

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isolating the microparticles from the water. In preferred embodiments, the organic solvent is chosen from the group of acetone, acetonitrile, tetrahydrofuran, dimethylformamide, and dimethoxy ethylene glycol.

5 In preferred embodiments the polyester/polypeptide ionic molecular conjugate is capable of releasing in vivo a therapeutically effective dose of a bioactive polypeptide over a period of at least 20 days, and more preferably for up to 95 days but not less than 7 days. In
10 yet other preferred embodiments the release of the therapeutic ionic molecular conjugate is essentially monophasic.

The sustained release compositions of the invention are preferably made by (a) providing a
15 polyester having free COOH groups and a bioactive polypeptide having at least one effective ionogenic amine, and (b) ionically conjugating the polyester to the polypeptide to form an ionic molecular conjugate, wherein at least 85%, by weight, of the polypeptide present in
20 the composition is ionically conjugated to the polyester. The polyester can be one which has sufficient free COOH groups to begin with, or, if an insufficient number of such groups are available for the desired peptide loading level at the outset, the polyester can be (1) reacted
25 with , e.g., malic or citric acid through esterification or functional interchange, or (2) acid-tipped with , e.g., glutaric anhydride or (3) the polyester can be treated with a base, e.g., NaOH, to expose acid groups. Finally, the polyester/polypeptide ionic molecular
30 conjugate may be converted to implantable films or rods, or injectable microspheres or microparticles capable of releasing in vivo the polypeptide.

Preferably, the polyester is synthesized by catalyzing or autocatalyzing direct condensation of one
35 or more hydroxy acids, e.g., glycolic acid and lactic

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acid, in the presence of a predetermined concentration of a polycarboxylic hydroxy acid, e.g., malic acid or citric acid. The polyesters thus formed possess acid-tipped hydroxyl end groups which are preferably partially
5 or fully acid-tipped.

The polyesters also can be synthesized by catalyzing ring-opening polymerization of lactones, or by polymerization of cyclic monomers such as ϵ -caprolactone, p-dioxanone, trimethylene carbonate, 1,5-dioxepan-2-one,
10 or, 1,4-dioxepan-2-one in the presence of a chain initiator, e.g., a hydroxy polycarboxylic acid.

Another method of synthesizing involves reacting a hydroxy acid with a cyclic dimer, followed by condensation of the open chain system in the presence of
15 a polycarboxylic acid.

Yet another synthetic method involves reacting an organic polycarboxylic acid with a preformed polyester.

In the aforementioned preferred embodiments the acid-tipped polyester has a carboxyl to hydroxyl end
20 group ratio of more than one and approaching infinity (i.e., eliminating all the hydroxyl groups) with an average degree of polymerization between 10 and 300, and in particularly preferred embodiments between 20 and 50.

Alternatively, a polyester is rendered capable of
25 forming an ionic molecular conjugate with a bioactive polypeptide by treatment with a base, e.g., NaOH.

Preferably, the polyester/polypeptide ionic molecular conjugate is synthesized by direct interaction between the polyester, e.g., in the free form, and the
30 polypeptide, e.g., in the free form, in the proper liquid medium. In other preferred embodiments suitable solvents for the formation of the conjugate would be a mixture aprotic solvent [e.g., acetone, tetrahydrofuran (THF), or ethylene glycol dimethylether] and a suitable solvent for
35 the peptide (e.g., water) in proportions such that the

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two systems are miscible. Preferably, the polypeptide is a salt of a monocarboxylic acid having a pKa greater than or equal to 3.5. Preferably, the polypeptide has at least one effective ionogenic amine group.

5 In preferred embodiments the polypeptide is 1 to 50 percent by weight, and preferably 10 to 20 percent of the polyester/polypeptide ionic molecular conjugate. In preferred embodiments, the accessible carboxyl groups of the polyester are partially neutralized with alkali metal
10 ions or organic bases. In yet other preferred embodiments, the alkali treatment provides chain dissociation of the polyester and formation of lower molecular weight binding sites.

"Polypeptide", as used herein, refers to a
15 protein, peptide, oligopeptide or synthetic oligopeptide.

"Polycarboxylic", as used herein, refers to compounds having more than one carboxyl group, e.g., malic acid and citric acid.

"Average degree of polymerization", as used
20 herein, refers to the number of repeat monomer sequences.

"Effective ionogenic amine", as used herein, refers to a polypeptide that contains at least one amine group capable of forming an ion under prevailing conditions.

25 "Acid-tipped", as used herein, refers to compounds having an acid terminus.

"Partially acid-tipped", as used herein, refers to compounds having 1 - 99 percent of their hydroxyl end groups acid-tipped.

30 "Fully acid-tipped", as used herein, refers to compounds having greater than 99.9% of their hydroxyl groups acid-tipped.

"Hydroxy acids", as used herein, refers to any compound containing hydroxyl and carboxyl groups.

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"Monocarboxylic hydroxy acid", as used herein, refers to an organic acid with one carboxyl group and one or more hydroxyl group.

"Polycarboxylic hydroxy acid", as used herein, refers to a hydroxy acid with more than one carboxyl group.

"Organic entrainer", as used herein, refers to organic liquids which codistill with water.

"Bioactive", as used herein, refers to a molecule that elicits or affects a biological event.

"Acyclize", as used herein, refers to a chemical reaction that occurs by ring opening.

"Polycondensation", as used herein, refers to the formation of a polyester by the condensation of two or more molecules.

The present invention provides a new pharmaceutical composition that chemically bonds a biocompatible, biodegradable polyester to oligopeptides, polypeptides, peptides and or proteins as a homogeneous ionic species. By chemically bonding polyesters of different molecular weights to the therapeutic agents, the chemical characteristics of the composition can be more precisely tailored to meet the demands for the controlled monophasic release of the biologically active polypeptide molecule in vivo. Furthermore, the compositions of the invention are optimized easily to possess functional properties for greater loading of a therapeutically active polypeptide.

Other features and advantages of the invention will be apparent from the following detailed description of the preferred embodiments, and from the claims.

Brief Description of the Drawings

Figure 1 is an illustration representing isomers of polycarboxylic acid-tipped lactide/glycolide (malic type) copolymer.

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Figure 2 is an illustration of an ionic molecular conjugate depicting the chemical interactions between the lactide/glycolide (malic type) copolymer and Somatuline (BIM-23014).

- 5 Figure 3 is a graph depicting the percent peptide released from ionic molecular conjugates into PBS buffer at 37°C over a 28-day period.

Description of the Preferred Embodiments

Synthesis

- 10 The biodegradable or absorbable polyesters of the invention are tailored to possess the desired chemical reactivity to provide controlled chain hydrolyzability and exhibit maximum binding capacity to oligopeptides, polypeptides or proteins having a net positive charge at
15 physiologic pH, by the proper selection of the constituent monomers, co-monomers or comers to form chains with predetermined compositions and molecular weights (see, e.g. Fig. 2).

- A tripartite synthetic design, within the ability
20 of someone of ordinary skill in this field, is employed to prepare the compositions of the present invention. The steps include: (1) synthesis of the polycarboxylic acid-tipped polyesters; (2) synthesis of the polyester/polypeptide ionic conjugate by ionic
25 interaction of polycarboxylic acid-tipped polyesters (or a polyester treated with a base) and biologically active polypeptides; and (3) conversion of ionic conjugates to implants, rods, microspheres or microparticles capable of releasing in vivo the therapeutic agent for at least 7
30 days.

1) Synthesis of polycarboxylic acid-tipped polyesters.

 The polycarboxylic acid-tipped polyester chains of the invention are synthesized by methods such as direct

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condensation of a 2-hydroxy acid and a polycarboxylic organic acid, step-growth polymerization of acyclized products, ring-opening polymerization of a lactone or a lactone mixture, or functional interchange of a

- 5 polycarboxylic organic acid with preformed high molecular weight polyesters (See Fig. 1) Descriptions of the synthesis of polycarboxylic acid-tipped polyesters by these aforementioned methods follow.

- Direct condensation of 2-hydroxy acids in an
- 10 optically active and/or an inactive form and a predetermined amount of a polycarboxylic organic acid in the presence or absence of inorganic, or organometallic catalyst, e.g., condensation of glycolic acid DL-lactic acid, and DL-malic acid is generally accomplished by
- 15 heating the monocarboxylic hydroxy acids or mixture of two or more monocarboxylic hydroxy acid in the presence of a fraction of polycarboxylic hydroxy acid in a glass reactor equipped for providing a continuous flow of dry-nitrogen and mass agitation (designated Type IA
- 20 Polyester, see Table I). Typically, the polycondensation is conducted at 150-170°C for 4 to 72 hours. Agitation of the reaction mixture can be provided by a magnetic stirrer or bubbling the nitrogen gas through the polyester mass. The polymerization is continued until
- 25 the desired average molecular weight (determined in terms of solution viscosity) and/or acid number (determined by end group titration) is achieved. Polyester analysis by end group titration is performed as follows. Polyester samples (300 mg-500 mg) are accurately weighed and
- 30 dissolved in a minimal amount (10-30 ml) of acetone. After dissolution, the solutions are diluted to 100 ml with benzyl alcohol (Mallinckrodt, Analytical Reagent) and titrated to a faint pink end-point (phenolphthalein) using potassium hydroxide in benzyl alcohol solution
- 35 (Normalized vs. HCl Standard). The volume of base

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solution used for the sample (ΔV_s) is compared with the volume of base used for a solvent blank (ΔV_o) to determine the acid number for the polyester.

$$\text{Acid \#} = \frac{\text{Wt. of Sample (mg)}}{\{\Delta V_s(\text{ml}) - \Delta V_o(\text{ml})\} \times N \text{ of Base}}$$

At the conclusion of polymerization, the polyester is isolated and extracted with water or a dilute aqueous sodium hydroxide solution, from a suitable organic solution to remove water-soluble or solubilizable low molecular weight chains.

Polyester analysis by GPC is performed as follows. The average molecular weights (MW) of polyester were determined by GPC using a Waters Model 6000 solvent delivery pump and a Dynamax (Rainin) model UV-D detector. Runs were performed in tetrahydrofuran (Burdick & Jackson UV grade) using a Jordin Gel DVB 1000Å, 50cm x 10 mm column (Jordi Associates) at a flow rate of 1.2 ml/min at 25 C. Peak detection was at 220 nm and 1.0 AUFS. The column was calibrated using narrow band polystyrene reference standards (Polysciences Inc.) at Mw = 4000, 9,200 and 25,000.

A modification of the direct condensation process entails the use of an organic entrainer and a cationic exchange resin as a condensation catalyst (designated Type IB Polyester, see Table I). This process requires a filtration and evaporation step to remove the catalyst and entrainer, respectively. Typical examples of polyesters made by these processes and pertinent analytical data are described in Table I.

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TABLE 1: POLYESTERS MADE BY DIRECT CONDENSATION METHOD

Type IA Polyesters						
Polymer #		Charge	Polymerization Conditions	Acid #	η_{inh}	T _g , °C
1	L-Lactic Acid (88%) Glycolic Acid Citric Acid	35.7 gm (0.349M) 4.65 gm (0.612M) 1.75 gm (0.0091M)	100°C/0.7 hr 165°C/17.5 hrs	563	0.24	11
2	L-Lactic Acid (88%) Glycolic Acid Malic Acid	25.6 gm (0.25M) 19.2 gm (0.25M) 1.5 gm (0.011M)	165°C/22 hrs	820	0.14	27
Type IB Polyesters						
3	L-Lactic Acid (88%) Glycolic Acid Citric Acid Amberlyst Catalyst Beads #15 Toluene	25.6 gm (0.25M) 19.2 gm (0.25M) 2.13 gm (0.011M) 0.5 gm 150 ml	132°C/53 hrs Using Dean-Stark Trap. Decanted, filtered in Acetone. Dried. Washed with water. Vacuum dried.	842	0.11	15
4	L-Lactic Acid (88%) Glycolic Acid Malic Acid Amberlyst Toluene	25.6 gm (0.25M) 19.2 gm (0.25M) 1.5 gm (0.011M) 100 ml	132°C/68 hrs Using Dean-Stark Trap. Decanted, Filtered, Dried. Washed with water and Vacuum Dried.	1421	0.20	28

*Determined on a differential scanning calorimeter (TA 2100 DSC) using
2-10 mg sample and heating rate of 10°C/min in a nitrogen atmosphere

Step-growth polymerization of acyclized products, where a hydroxy acid is allowed to react with cyclic dimers and subsequent condensation of the resulting open chain system in the presence of predetermined amounts of a polycarboxylic acid and the presence or absence of a suitable condensation catalyst e.g. glycolic acid, L-lactide and DL-malic acid, essentially the same as the condensation process described above, except that it employs a mixture of a monocarboxylic hydroxy-acid, a cyclic dimer of a second

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hydroxy-acid, and a hydroxy polycarboxylic acid.

Examples of polyesters made by this process and pertinent analytical data are summarized in Table II. When the cyclic dimer is pretreated with water the system is

5 treated as a simple step-growth polymerization.

TABLE II: STEP-GROWTH POLYMERIZATION OF ACYCLIZED PRODUCTS

Type II Polyesters							
Polymer #		Charge		Polymerization Conditions	Acid #	η_{inh}	T _g , °C*
1	L-Lactide Monomer	10.0 gm	(0.07M)	160°C/29 hrs	1200	0.21	20
	Glycolic Acid	10.7 gm	(0.14M)	Malic Acid			
		0.79 gm	(0.0061M)				
2	L-Lactide Monomer	20.0 gm	(0.139M)	25°C-155°C/1.5h	1800	0.13	27
	Glycolic Acid	7.1 gm	(0.093M)	155°C/70 hr			
	Malic Acid	1.01 gm	(0.0075M)	Dissolve in DCM wash with water, and dry <u>in vacuo</u> .			

* Determined on a differential scanning calorimeter (TA2100 DSC) using 2-10 mg sample and heating rate of 10°C/min in a nitrogen atmosphere

Ring opening polymerization of a lactone or lactone mixture in the presence of a predetermined concentration of hydroxy-polycarboxylic acid as a chain initiator and a catalytic amount of an organometallic catalyst e.g. a mixture of L-lactide, glycolide and DL-malic acid in the presence of stannous octoate employs dry cyclic monomers or a mixture of cyclic monomers, a hydroxy-polycarboxylic acid and a trace amount of stannous octoate (used as 0.33M solution in toluene), which are transferred under a dry oxygen-free atmosphere into a glass reactor equipped for magnetic or mechanical stirring. The polymerization reaction is continued under nitrogen following a suitable heating scheme until the desired molecular weight is achieved (as measured in terms of solution viscosity). At the conclusion of the polymerization scheme, the temperature is lowered and the unreacted monomer is distilled under reduced pressure. The polyester mass is then cooled and the water-soluble

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low molecular weight fractions are removed by low temperature extraction from a suitable organic solution. The solution is then dried and the solvent is removed. The molecular weight is then determined in terms of inherent viscosity and the acid number is determined by end-group titration. Examples of polyesters prepared by this process and pertinent analytical data are given in Table III.

TABLE III: POLYESTERS MADE BY RING OPENING POLYMERIZATION

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TABLE III. POLYESTERS MADE BY KRIC OPENING POLYMERIZATION

Type III Polyesters							
Polymer #	Charge	Polymerization Conditions	Acid #	η_{inh}	$T_g, ^\circ C^*$		
1	Glycolide 3.22 gm (0.028M) L-Lactide 10.7 gm (0.14M) Malic Acid 0.79 gm (0.0061M)	120°C/0.5 hr 150°C/6 hrs 120°C/11 hrs	2,150	0.79	38		
2	Glycolide 2.84 gm (0.0245M) D,L-Lactide 20.0 gm (0.139M) Malic Acid 0.876 gm (0.00541M)	120°C/0.5 hr 180°C/2.5 hrs 130°C/15 hrs	1,206	0.08	26		
3	Glycolide 2.84 gm (0.0245M) D,L-Lactide 20.0 gm (0.139M) Citric Acid 1.256 gm (0.00654M)	155°C/1 hr 185°C/2.5 hrs 190°C/2.5 hrs 160°C/13 hrs	937	0.10	27		
4	Glycolide 8.06 gm (0.0694M) D,L-Lactide 10.0 gm (0.0694M) Malic Acid 0.744 gm (0.00555M)	180°C/1 hr 185°C/2 hrs 195°C/7 hrs 120°C/9 hrs	970	0.26	23		
5	Glycolide 8.06 gm (0.0694M) D,L-Lactide 10.0 gm (0.0694M) 1,6-hexanediol 0.656 gm (0.00555M)	150°C/0.5 hr 185°C/4 hrs 150°C/1.5 hrs 120°C/3 hrs	10,138	0.39	30		

* Determined on a differential scanning calorimeter (TA2100 DSC) using 2-10 mg sample and heating rate of 10°C/min in a nitrogen atmosphere

** In hexafluoroisopropanol

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Functional interchange of polycarboxylic or hydroxy-polybasic organic acid with preformed high molecular weight polyesters with COOH/OH ratio of unity to virtually zero, preferably in the presence of an organometallic catalyst, e.g. melt-reaction of an 85/15 lactide/glycolide copolymer with a molecular weight greater than 5,000 and COOH/OH ≤ 1 with DL-malic acid in the presence of stannous octoate, to produce the lower

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molecular weight polyesters with $\text{COOH}/\text{OH} \geq 1$, entails heating a high molecular weight polyester with a predetermined amount of polycarboxylic or hydroxy-polycarboxylic acid in the presence of a trace amount an
5 organometallic catalyst such as stannous octoate. The reactants are heated above 150°C under dry nitrogen with intensive agitation until the functional interchange is completed (as measured by depletion of residual unreacted polycarboxylic acid). In effect this is determined by
10 monitoring the molecular weight (in terms of solution viscosity using capillary viscometry at 28°C) of the resultant lower molecular weight polyester and the presence of unreacted polycarboxylic acid. This is achieved by aqueous extraction of a polyester sample and
15 analysis of the extract using high performance liquid chromatography (HPLC). Residual monomer, dimer and polycarboxylic acid levels were determined by HPLC using a Waters Model 6000 solvent delivery pump and a Dynamax (Rainin) model UV-D detector (205 nm, 1.0 AUFS). Runs
20 were performed using 0.025N Na_2PO_4 buffer, pH = 3.5 (isocratic flow rate = 1.0 ml/min.) using a Nucleosil C18, 5 μm , 25 cm x 4.6 mm column.

The desired polyester is isolated and purified as described above for ring-opening polymerization. An
25 example of a polyester made by this process and pertinent analytical data are given in Table IV.

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TABLE IV: POLYESTERS MADE BY FUNCTIONAL INTERCHANGE

Type IV Polyesters						
5	Polymer #	Charge	Polymerization Conditions	Acid #	η_{inh}	T _g , °C*
	1	Boehringer A001 8 gm (50/50 dl-Lactide/Glycolide)	150°C/5 hrs	670	0.26	25
		Citric Acid** 0.8 gm (0.00417M)				
10	* Determined on a differential scanning calorimeter (TA2100 DSC) using 2-10 mg sample and heating rate of 10°C/min in a nitrogen atmosphere					
	** A catalytic amount of stannous octoate (2 drops of 0.33 M solution, approximately 0.03 nmole).					

- 15 Among the other monomers suitable for the synthesis of the polyesters used in the invention are: L-lactic acid, DL-lactic acid, ϵ -caprolactone, p-dioxanone, ϵ -caproic acid, trimethylene carbonate, 1,5-dioxepan-2-one, 1,4-dioxepan-2-one, glycolide, and meso-lactide.
- 20 Examples of a useful polycarboxylic chain initiators and/or chain modifiers include malic acid, and citric acid.

- 25 (2) Synthesis of the polyester/polypeptide ionic conjugate by ionic interaction of polycarboxylic acid-tipped polyesters and biologically active polypeptides.
- The polycarboxylic acid-tipped biodegradable polyesters described above are used to make an ionic molecular conjugates with mono- or polycarboxylic oligopeptides, polypeptides or proteins with accessible effective
- 30 ionogenic amine groups (see Fig. 2). Moreover, any polyester is rendered capable of forming an ionic molecular conjugate with a polypeptide provided it is treated with a base, e.g., 0.1N NaOH. Such treatment exposes acid groups of the polyester for multiple site
- 35 ionic interaction with the cationic polypeptide.

Thus, the formation of these conjugates is achieved by direct molecular interaction of the components in the proper solvent with or without a pretreatment of the polyester with an inorganic base to

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maximize its binding rate capacity to the basic drug. As noted above, the ionic interaction of their ionic conjugate components increases within the difference in their pKa values.

5 The polyester is dissolved in a suitable aprotic solvent in a concentration range of 2% to 20% W/V. Such solvents should dissolve the polyesters, but also be partly miscible with water. Suitable solvents used for
10 ethylene glycol dimethylether. To this solution, an aqueous solution of base such as sodium, potassium or ammonium hydroxide or carbonate is added to maximize the binding capacity of the polyester. In general, the amount of base added corresponds to the amount of acid
15 represented by the counter-anion level of the basic peptide to be used.

After briefly mixing the polyester-base combination, an aqueous solution of peptide or peptide salt is added at peptide/polyester loading levels of 2%
20 to 50% W/W (peptide/polyester). This mixture is stirred for a period of time (up to 3 hours), and then the solvents are removed and the product is dried under vacuum. The resulting material may then be further processed for dosage formulation. The resulting
25 pharmaceutical compositions are designed to be chemically uniform compositions made totally of ionic molecular conjugates and are essentially devoid of microscopically or macroscopically dispersed domains of the active drug in the biodegradable matrix. Examples of
30 prepared ionic molecular conjugates and pertinent analytical data are given in Table V.

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TABLE V: IONIC MOLECULAR CONJUGATE-PEPTIDE BINDING¹

	Polymer Used	Peptide ²	Loading %	Retention ³ %
5	1 50/50 dl Lactide/Glycolide (Commercial) Acid # = 22,000 η_{inh} = 0.53	I I II III	10 20 20 20	47 25 73 48.5
10	2 Poly L-Lactide (Commercial) Mw (ave) = 2,000 Acid # = 850	I II	10 20	62 40
15	3 Poly L-Lactide (Commercial) Mw (ave) = 50,000 Acid # = 2100	I	10	54
20	4 48/48/4 Poly d,l-Lactide/ Glycolide/1,6 hexanediol (Method III) Acid # = 10,138 η_{inh} = 0.39	I	20	43
25	5 49/49/2 Poly L-Lactic/ Glycolic/Malic Acid (Type IB) Acid # = 1400 η_{inh} = 0.20	I I I I I II III	10 20 30 40 50 20 20	100 99 95.5 96.0 99.8 99.8 77.5
30	6 83.3/14.7/2 Poly L-Lactic/ Glycolic/Citric Acid (Type IA) Acid # = 563 η_{inh} = 0.24	I	20	96
35	7 49/49/2 Poly d,l-Lactide/ Glycolide/Malic Acid (Type II) Acid # = 1200 η_{inh} = 0.21	I III	20 20	96 73.9
40	8 48/48/4 Poly d,l-Lactide/ Glycolide/Citric Acid (Type III) Acid # = 589 η_{inh} = 0.22	I	10	90

45 ¹ In all cases, conjugates were formed as outlined in text using acetone as solvent and sodium hydroxide as base. All peptides used were in the acetate salt form.

² Peptides: I BIM-21003 D-Trp⁶-LHRH(pGlu-His-Trp-Ser-Tyr-D-Trp-Leu-Arg-Pro-Gly NH₂) pKa = 10.1

50 II BIM-23014 (H₂N-β-D-Nal-Cys-Tyr-Trp-Lys-Val-Cys-Thr NH₂) pKa = 9.8

III BIM-26226 (H₂N-D-F₅ Phe-Gln-Trp-Ala-Val-D-Ala-His-Leu-OCH₃) pKa = 8.0

³ % Retention: Measured by rinsing the dried polyester/peptide ionic conjugates with D.I. water and quantitating soluble peptide in rinses by HPLC

55 % Retention = 100% x $\frac{\text{Wt. Peptide Loaded} - \text{Wt. Soluble Peptide}}{\text{Wt. Peptide Loaded}}$

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(3) Conversion of ionic conjugates to implants, rods, microspheres or microparticles capable of releasing in vivo the therapeutic agent for at least 20 days in a monophasic profile. The ionic conjugate salts of the invention can be

- 5 converted to: (A) sterile injectable microspheres (with or without 0.1 to 10% of a solid polyhydric alcohol as processing aid) containing 1 to 50% by weight polypeptide that can released according to an essentially monophasic profile and to sustain pharmacological activity over a
10 period of one to 12 weeks; (B) sterile implantable films made by casting, pressing or extrusion with or without a pharmacologically inactive processing aid, and capable of providing a release profile similar to that described in (A) and; (C) sterile injectable rods made by extrusion or
15 pressing, capable of providing a release profile similar to that described in (A).

In-Vitro Release Assay:

- Samples of the dried and ground ionic conjugate material weighing 50 mg each were placed in 25 mm diameter
20 scintillation vials A 5 ml aliquot of modified PBS buffer (PBS buffer: 2.87 gm Na_2HPO_4 , 0.654 gm NaH_2PO_4 , 5.9 gm NaCl , 0.5 gm NaN_3 , Q.S. 1.0 Liter with Deionized water; pH = 7.27.) was added to each vial and the vials were placed in a Lab-Line Orbit Environ-Shaker and swirled at 120
25 R.P.M. and 37°C. Vials were withdrawn periodically and decanted and replenished with fresh PBS solution. The amount of released peptide was determined from the decanted PBS solutions by HPLC.

Peptide Extraction From Ionic Conjugates:

- 30 A 50 mg sample of an ionic molecular conjugate was mixed into 20 mls of methylene chloride. The mixture was sequentially extracted with 50 ml, 20 ml and 20 ml portions of 2N acetic acid. The acetic acid extracts were combined and analyzed for peptide content by high performance liquid
35 chromatography (HPLC). Peptide analysis by HPLC is as follows. HPLC analysis were performed using a Waters model M-45 solvent delivery pump and an EM Science MACS 700

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detector at wavelength 220 nm and 1.0 AUFS. Peptides were run using a Lichrospher (EM separations) C18, 100Å, 5µm, 25cm x 4.6 mm column and 30% acetonitrile/0.1% TFA as an isocratic eluent buffer.

5 Following are details (Table VI) of the in vitro assay demonstrating the amount of peptide released over a 28 day period for 49:49:2 L-Lactic/Glycolic/Malic\D-Trp⁶[LHRH] (Example #8), 49:49:2:L-Lactic/Glycolic/Malic\Somatostatin-Tumor Inhibiting Analogue (Example #9), and 73.5:24.5:2
10 poly-L-lactide/glycolic/Malic:D-Trp⁶[LHRH] (Example #10) ionic molecular conjugates. Fig. 3 shows the graphic representation of these data.

TABLE VI IN-VITRO ASSAY DATA

15 DAY OF ASSAY	PERCENT OF TOTAL PEPTIDE RELEASED		
	<u>Example #8</u>	<u>Example #9</u>	<u>Example #10</u>
1	5.5%	12.5%	11%
7	26.9%	21.3%	53%
14	55.2%	47.3%	55%
20 17	84.4%	72.2%	60%
21	98.6%	82.5%	66%
24	100%	98.2%	75%
28	---	99.6%	---

25 Quantitation of Peptides in Ionic Conjugates

Ionically bound peptides in the conjugate products were measured by dissolving 10 mg samples in 5.7 ml of a 9:1 mixture of acetone and 0.1M aqueous trifluoroacetic acid. The solutions were swirled at 25° for 15-24 hours and then
30 filtered through 0.5 µm teflon filter cartridges. The filtrates were then analyzed for peptide content by high performance liquid chromatography (HPLC). Peptide analysis by HPLC were run using a Millipore model 717 Wisp Autosampler, a model 510 pump and a model 486 UV
35 detector set at 220nm. Peptides were run on a Lichrospher (EM Separations) 25 cm x 4.6 mm C18, 5 µm 100Å column, flow rate 1.0 ml per minute using 35% acetonitrile in 0.14% sodium perchlorate buffer as an isocratic eluent system. Peptides were quantified by

vacuum drying said rinsed conjugate.

28. A method according to claim 27, wherein said isolated conjugate is mixed with an aqueous mannitol solution prior to vacuum drying.

5 29. A method of spherifying a sustained release ionic conjugate containing a free carboxyl group-containing biodegradable polymer and a free amino group-containing drug which are ionically bonded to each other, said method comprising:

dissolving said conjugate in a first liquid to form a first solution;

10 stirring said first solution with a second liquid to form a first dispersion, wherein said second liquid is immiscible with said first solution;

evaporating said first liquid from said first dispersion to precipitate said conjugate from said first dispersion; and

15 isolating said precipitate conjugate from said first dispersion.

30. A method according to claim 29, wherein said first solution is added to said second liquid as small droplets.

31. A method according to claim 29 or 30, wherein said first liquid is acetonitrile and said second liquid is an oil.

20 32. A method according to claim 31, wherein said oil is silicon oil, mineral oil, sesame oil, or a vegetable oil.

33. A method according to any one of claims 29-32, wherein said biodegradable polymer is a polyester comprising lactic acid or glycolic acid; or a copolymer thereof.

34. A method according to any one of claims 29-33, wherein said drug is a peptide.
35. A method according to any one of claims 29-34, further comprising rinsing said isolated conjugate with a third liquid which is
5 miscible with said second liquid and not a solvent for said isolated conjugate.
36. A method according to claim 35, wherein said third liquid is hexane, heptane, or octane.
37. A method according to any one of claims 29-36, wherein
10 said isolated conjugate is mixed with an aqueous mannitol solution prior to vacuum drying.
38. A method of making microparticles of a sustained release ionic conjugate, substantially as hereinbefore described and exemplified.
39. A method of spherifying a sustained release ionic
15 conjugate, substantially as hereinbefore described and exemplified.

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70 hours the reaction was cooled to 100°C and poured into a chilled stainless steel receiver for hardening. The solid polyester was then dissolved in methylene chloride and was washed twice with water (2 x 150 ml) to remove soluble oligomers. The organic solution was concentrated on a rotary evaporator and the product was thoroughly dried under vacuum to afford a white solid (see Table II, Type II Polyester, Polymer #2).

(η_{inh} in $CHCl_3$ = 0.13, Acid # = 1800, T_g = 27°C)

10 **EXAMPLE 4 -- RING OPENING POLYMERIZATION METHOD --**
Synthesis of 75/25 poly(L-lactide-co-glycolide) initiated by malic acid.

L-lactide (12.0 g, 0.0833 mole), glycolide (3.21 g, 0.0277 mole), malic acid 0.3042 g, 0.00227 mole), and
15 stannous octoate catalyst (0.33 M in toluene, 67 μ L, 0.022 mmole) were added under dry nitrogen conditions to a glass ampule with a magnetic stirrer. The system was purged with N_2 and evacuated with vacuum several times before sealing the ampule. The reactants were then
20 melted at 140°C and the melt was heated at 180°, 190°, 180°, and 150° for 1, 4.5, 12, and 2 hours, respectively. After cooling to room temperature, the polyester was reheated to 110°C under a vacuum of less than 1 mm Hg for about one hour to remove monomer, recooled at room
25 temperature, quenched in liquid nitrogen, isolated and dried under vacuum

(η_{inh} in $CHCl_3$ = 0.20, Acid # = 2560, T_g = 39°C)

EXAMPLE 5 --RING OPENING POLYMERIZATION METHOD--
Synthesis of 50/50 poly(D,L-lactide-co-glycolide)
30 **initiated by citric acid.**

D,L-lactide (10.0 g, 0.0694 mole) was mixed with glycolide (8.06 g, 0.0694 mole), citric acid (1.07 g, 0.00555 mole) and stannous octoate catalyst (0.33 M in toluene, 84 μ L, 0.0278 mmole) under a dry nitrogen
35 atmosphere in a glass ampule containing a magnetic stirrer

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and sealed under vacuum. The reactants were melted and heated at 180°, 185°, 195°, and 120°C for 1, 2, 7, and 9 hours, respectively. The polyester was cooled to room temperature, quenched in liquid nitrogen, isolated, and
5 dried.

(η_{inh} in $CHCl_3$ = 0.26, Acid # = 970, T_g = 23°C)

EXAMPLE 6 --RING OPENING POLYMERIZATION METHOD -- Synthesis of 50/50 poly(D,L-lactide-co-glycolide) initiated by 1,6-hexanediol.

10 Using a system similar to those described above, D,L-lactide (10.0 g, 0.0694 mole), glycolide (8.06 g, 0.0694 mole), 1,6-hexanediol (0.656 g, 0.00555 mole), and stannous octoate (0.33 M in toluene, 84 μ L, 0.0278 mmole) were added under dry nitrogen conditions to a glass ampule which was
15 subsequently sealed under vacuum. The constituents were heated at 150°, 185°, 150°, and 120°C for 0.5, 4, 1, 5, and 3 hours, respectfully. The resulting polyester was recovered and dried (see Table III, Type III Polyester, Polymer #5).

20 (η_{inh} in $CHCl_3$ = 0.39, Acid # = 10,138, T_g = 30°C)

EXAMPLE 7 -- FUNCTIONAL INTERCHANGE METHOD - Synthesis of carboxylic-bearing 50/50 poly(D,L-lactide-co-glycolide).

50/50 poly(D,L-lactide-co-glycolide) (Boehringer A001, 8 g), citric acid (0.8 g, 4.16 mmole), and stannous octoate
25 (2 drops) were added to a glass ampule under dry nitrogen conditions and sealed. The mixture was heated at 150° for 4 hours, cooled to room temperature, quenched in liquid nitrogen, isolated, and dried (see Table IV, Type IV Polyester, Polymer #1).

30 (η_{inh} in $CHCl_3$ = 0.26, Acid # = 670, T_g = 23 C)

EXAMPLE 8 -- Synthesis of a 49:49:2 L-Lactic/Glycolic/Malic (See Table I, Polymer #4) and D-Trp⁶ [LHRH] ionic molecular conjugate.

- 25 -

500 mg of 49:49:2 L-Lactic/Glycolic/Malic (synthesized by direct condensation; Mw = 9,500; Acid# = 1420) was dissolved in 10 mls of Acetone (Mallinckrodt Analytic Reagent). A portion of 0.1N sodium hydroxide solution (1.14 ml) was added and the mixture was stirred at room temperature for 15 minutes. A solution of 100 mg of D-Trp⁶ [LHRH] (BIM-21003 Peptide I; base content 87%, acetate content 7%) in 1.0 ml water was added and the mixture was stirred for 1 hour at room temperature. The solvents were then removed, first by Rotovap at T<40°C and then in a desiccator for 1 hour at room temperature under 1 mm Hg vacuum. The dried solid was triturated and stirred in 100 mls of deionized Water and isolated by filtration. The aqueous filtrate was tested by HPLC and found to contain <1 mg soluble peptide. The solid material was dried several days in vacuo to give 540 mg of white powder. The powder was used in an in vitro assay (see Table VI, Example #8).

EXAMPLE 9-- Synthesis of a 49:49:2 L-lactic/glycolic/malic, see Table I, polymer #4) and Somatostatin/Tumor Inhibiting Analogue ionic molecular conjugate.

100 mg of 49:49:2 L-Lactic/Glycolic/Malic (synthesized by direct condensation; Mw = 9,500; Acid # = 1420) was dissolved in 2mls of acetone (Mallinckrodt Analytic Reagent). A portion of 0.1N sodium hydroxide solution (0.32 ml) was added and the mixture was stirred at room temperature for 15 minutes. A solution of 20 mg of Somatostatin/Tumor Inhibiting Analogue (BIM-23014 Peptide II; base content 83%, acetate content 9.8%) in 1.2 ml water was added and the mixture was stirred for 1 hour at room temperature. The solvents were then removed, first by Rotovap at T<40°C and then in a desiccator for 1 hour at room temperature under 1 mm Hg vacuum. The dried solid was triturated and stirred in 20 mls of Deionized Water and isolated by filtration. The aqueous filtrate was tested by HPLC and found to contain <0.05 mg soluble peptide. The solid material was dried several days in vacuo to give 106

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mg of white powder. The powder was ground and used in an in vitro release assay (see Table VI, Example #9).

EXAMPLE 10 -- Synthesis of a 73.5:24.5:2 poly L-lactide/glycolic/malic (see Table II, see polymer #2) and D-Trp⁶ [LHRH] ionic molecular conjugate.

800 mg of 73.5:24.5:2 poly L-lactide/glycolic/malic (synthesized by step growth of acylized products; Acid# = 1800) was dissolved in acetone (16 ml). A portion of 0.1N sodium hydroxide solution (2.8 ml) was added and the solution was stirred at room temperature for 20 minutes. A solution of 200 mg of D-Trp⁶ [LHRH] (BIM-21003; base content 87%, acetate content 7%) in 2 ml water was added and the mixture was stirred for 90 minutes. The solvents were removed and the resulting solid was triturated in deionized water as in Example 8 indicating less than 1% soluble peptide salt present. The isolated solids were dried 4 days in vacuo to give 839 mg of white powder. The powder was ground and used for in vitro release assay (see Table VI, Example #10).

EXAMPLE 11 -- Formation of Peptide-polymer Ionic Conjugate Microparticle 1.50 of L-lactide/glycolide/d,l malic acid polyester (65; 33; 2).

Conjugates were synthesized by ring opening polymerization as in example 4 (MW = 4700 polydispersity = 1.3 as determined by GPC on a Jordi Gel 50 x 1 cm mixed linear bed column, THF eluent, Wyatt Mini Dawn light scattering detector dn/dc = 0.05, acid #1475 by titration, T_g = 42°C], was dissolved in 40 ml of acetone. The acid groups were neutralized with 2.0 ml of 0.5 g of BIM-23014 (peptide content 83.7%, acetate content 11.5%) in 20 ml of Milli-Q water was slowly added with mixing to the polymer solution. An additional 40 ml of acetone was also added portionwise during the peptide addition to prevent precipitation. The clear, colorless solution was stirred for one hour and then evaporated to dryness under vacuum.

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The resulting white solid was redissolved in a mixture of 20 ml of acetone and 2 ml of Milli-Q water to form a clear solution. This solution was injected, via a 0.2 μ teflon filter into a rapidly stirring reservoir of 500 ml Milli-Q water at 4°C. The polymer/peptide complex phase separated immediately into small particles upon contact with the water. After mixing the slurry for 30 min at 4°, the residual acetone was removed under reduced pressure and the solids were isolated by centrifugation, resuspended with 100 ml of Milli-Q water, and recentrifuged. The isolated solids were dried by lyophilization to give 1530 mg of white free flowing powders. Particles size range = 2 - 100 μ m. The T_g of the ionic conjugate was shown to occur at 53°C. Total residual (unbound) peptide in all aqueous supernatants was found to be 63 mg by HPLC analysis. The total initial peptide content was determined by elemental nitrogen analysis to be 19.9% by weight. The percentage of extractable peptide from the conjugate was determined to be 16.9%, by weight, using the acetone/0.1M TFA extraction technique. The resulting conjugate, thus, retains 84.8% ionic (extractable) character.

From the foregoing description, one skilled in the art can easily ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions. Thus, other embodiments are also within the claims.

What is claimed is:

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Claims

1. A composition comprising a polyester containing one or more free COOH groups ionically conjugated with a bioactive polypeptide comprising at least one effective ionogenic amine, wherein at least 50%, by weight, of the polypeptide present in said composition is ionically conjugated to said polyester.
2. The composition of claim 1, wherein said polyester has a carboxyl to hydroxyl ratio greater than one.
3. The composition of claim 1, wherein said polyester is composed of a member chosen from the group of L-lactic acid; D-lactic acid; DL-lactic acid; ϵ -caprolactone; p-dioxanone; ϵ -caproic acid; alkylene oxalate; cycloalkylene oxalate; alkylene succinate; β -hydroxybutyrate; substituted or unsubstituted trimethylene carbonate; 1,5-dioxepan-2-one; 1,4-dioxepan-2-one; glycolide; glycolic acid; L-lactide; D-lactide; DL-lactide; meso-lactide; and any optically active isomers; racemates or copolymers thereof.
4. The composition of claim 1, wherein said polyester is partially acid-tipped with glutaric anhydride.
5. The composition of claim 1, wherein said polyester is fully acid-tipped with glutaric anhydride.
6. The composition of claim 1, wherein said polyester has an average degree of polymerization between 10 and 300.

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7. The composition of claim 1, wherein said polyester has a viscosity of about 0.05 - to about 0.7 dl/g in chloroform and an average molecular weight of about 1200 - 40,000.

5 8. The composition of claim 1, wherein said bioactive polypeptide comprises 1 to 50 percent of total weight of said ionic molecular conjugate.

9. The composition of claim 1, wherein greater than 85% of said polypeptide present in said composition is
10 ionically conjugated to said polyester.

10. The composition of claim 1, wherein said polypeptide is chosen from the group consisting of LHRH, somatostatin, bombesin/GRP, calcitonin, bradykinin, galanin, MSH, GRF, amylin, tachykinins, secretin, PTH,
15 CGRP, neuromedins, PTHrP, glucagon, neurotensin, ACTH, GHRP, GLP, VIP, PACAP, enkaphelin, PYY, motilin, substance P, NPY, TSH, and analogs or fragments thereof.

11. The composition of claim 1, wherein said ionic conjugate is capable of releasing in vivo a
20 therapeutically effective dose of said polypeptide over a period of at least 7 days.

12. A method of synthesizing a composition comprising (a) providing a polyester and a bioactive polypeptide having at least one effective ionogenic
25 amine, and (b) ionically conjugating said polyester to said polypeptide to form an ionic molecular conjugate in which the polypeptide is ionically conjugated to the polyester.

13. The method of claim 12, wherein at least 50%,
30 by weight, of the polypeptide present in the composition is ionically conjugated to the polyester.

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14. The method of claim 13, wherein said polyesters possess acid-tipped hydroxyl end groups.

15. The method of claim 14, wherein said hydroxyl end groups are partially acid-tipped with glutaric anhydride.

16. The method of claim 14, wherein said hydroxyl end groups are fully acid-tipped with glutaric anhydride.

17. The method of claim 13, wherein said polyesters are synthesized using hydroxy polycarboxylic acids chain initiators.

18. The method of claim 13, wherein the hydroxyl end-groups of said polyesters are acid-tipped.

19. The method of claim 18, wherein said hydroxyl end-groups are partially acid-tipped with glutaric anhydride.

20. The method of claim 18, wherein said hydroxyl end-groups are fully acid-tipped with glutaric anhydride.

21. The method of claim 13, wherein the synthesis of said polyester results in an average degree of polymerization between 10 and 300.

22. The method of claim 21, wherein said polyester has a carboxyl to hydroxyl end group ratio greater than one.

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23. The method of claim 13, wherein synthesizing said polyester/polypeptide ionic molecular conjugate comprises: (a) dissolving the polyester in tetrahydrofuran, acetone, or ethylene glycol dimethylether followed by adding a base; and (b) adding an aqueous solution of a polypeptide or salt of a polypeptide at polypeptide/polyester loading levels of 2% to 50% w/w (polypeptide/polyester).

24. The method of claim 23, wherein said polypeptide is a salt of an acid having a pKa greater than or equal to 3.5

25. The method of claim 23, wherein said polypeptide comprises 1 to 50 percent by total weight of said ionic conjugate.

26. The method of claim 23, wherein greater than 85% of the said polypeptide present in said composition is ionically conjugated to said polyester.

27. The method of claim 23, wherein said reaction results in the formation of an ionic bond between said reactants.

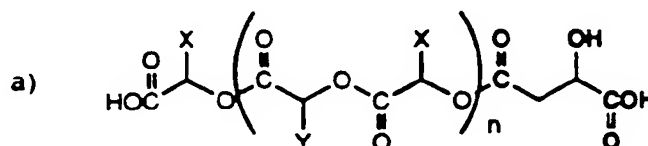
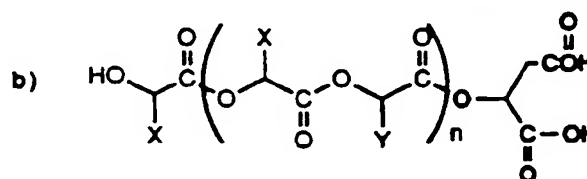
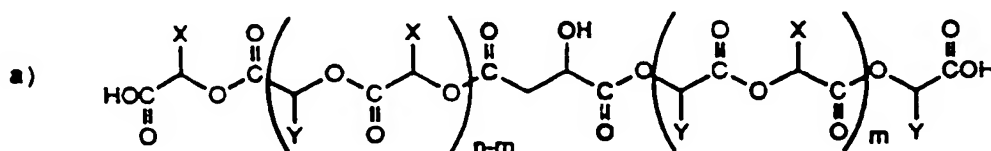
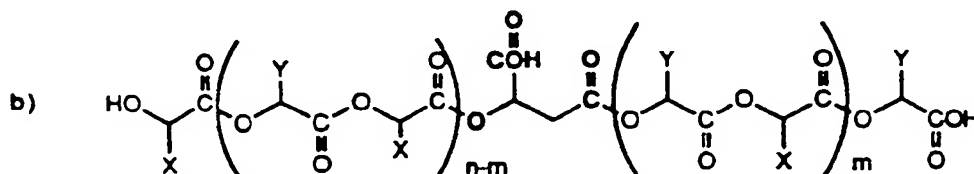
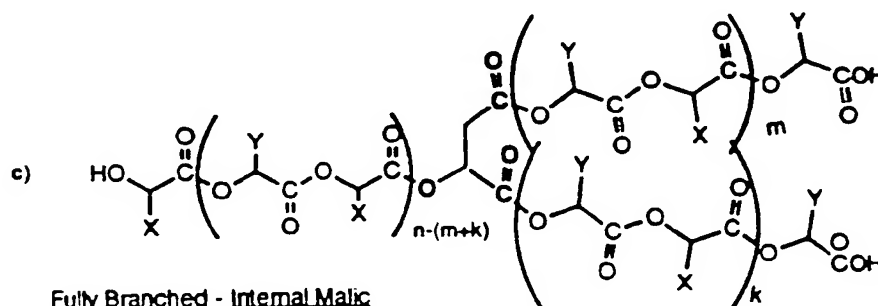
28. A composition made by the process of claim 12.

29. A method of synthesizing a microparticle comprising (a) dissolving the composition of claim 1 in an aprotic, water miscible organic solvent; (b) mixing said organic solvent in water; and (c) isolating said microparticles from said water.

30. The method of claim 29, wherein said organic solvent is chosen from the group consisting of acetone, acetonitrile, tetrahydrofuran, dimethylformamide, and dimethoxy ethylene glycol.

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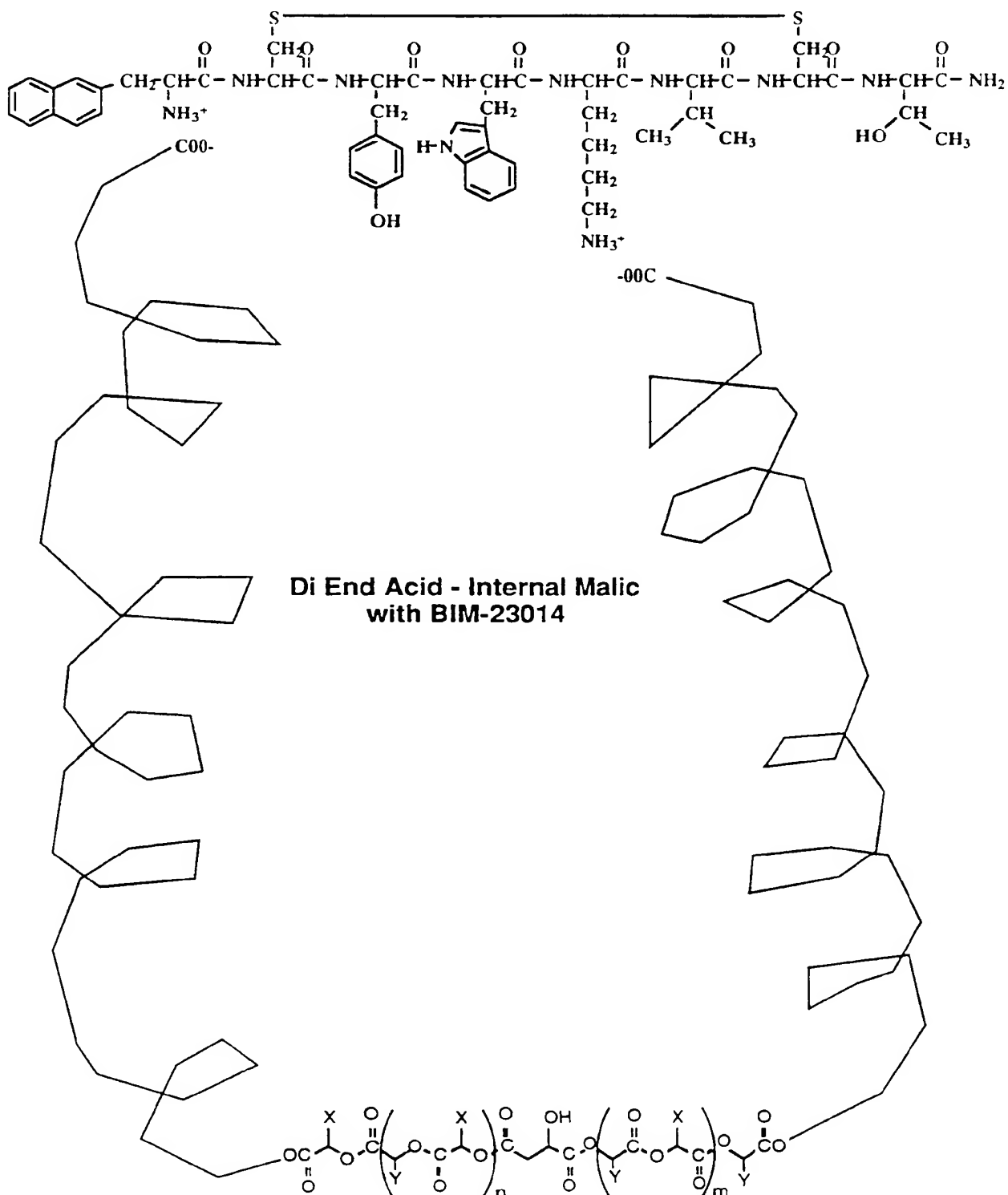
Forms of Lactate / Glycolate - Type Polymers of Polycarboxylic Acid-Tipped Chains

Di End Acid - Terminal Malic**I. Linear Chains**End Hydroxy - Terminal Malic**II. Branched Chains**Di End Acid - Internal MalicEnd Hydroxy - Internal MalicFully Branched - Internal Malic

Y = CH₃ for Lactic, H for Glycolic; X = H for Glycolic, CH₃ for Lactic
Distribution of X + Y is formula dependent and polymerization dependant

FIG. 1**SUBSTITUTE SHEET (RULE 26)**

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IONIC CONJUGATE EXAMPLE**FIG. 2****SUBSTITUTE SHEET (RULE 26)**

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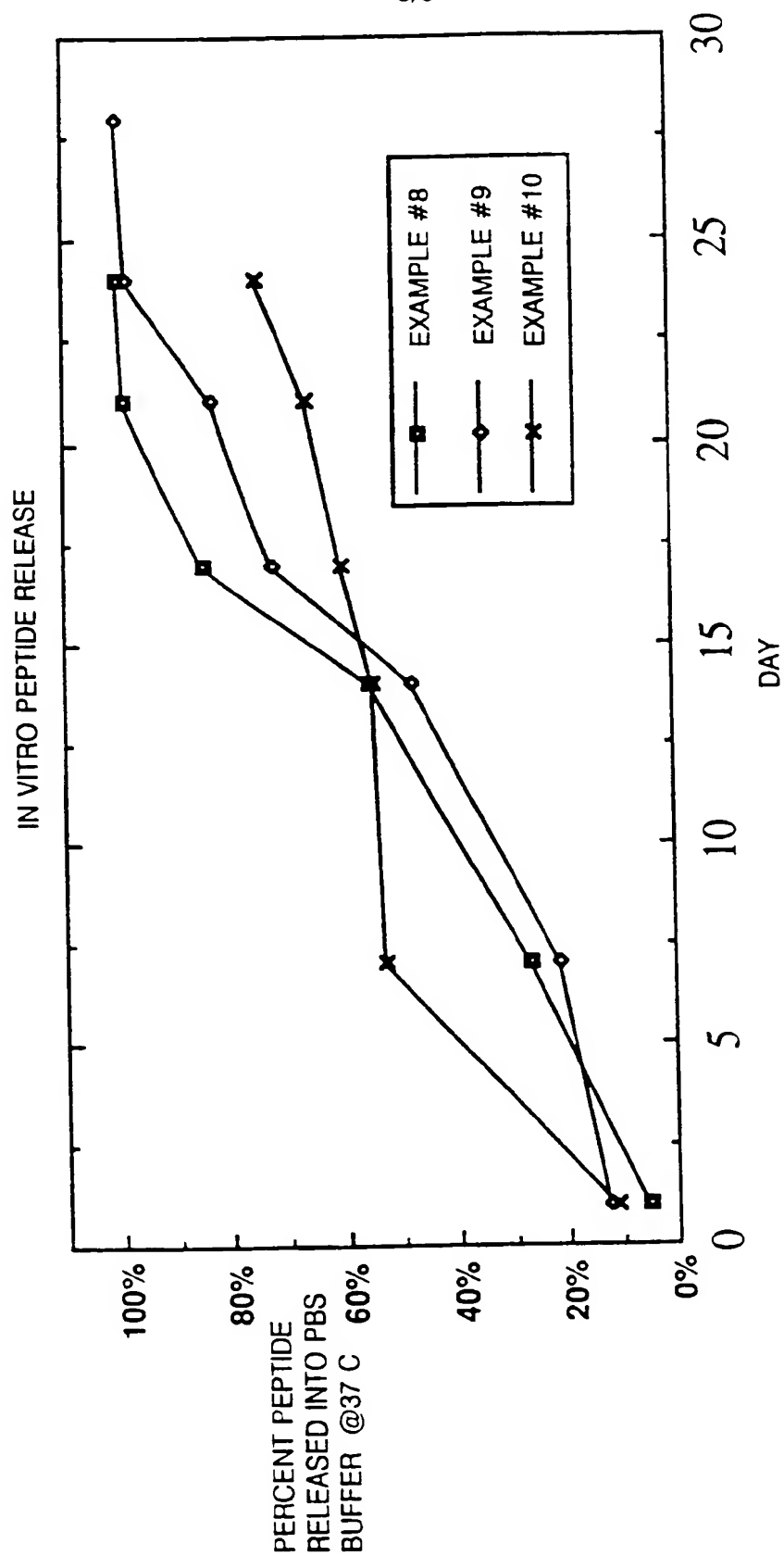
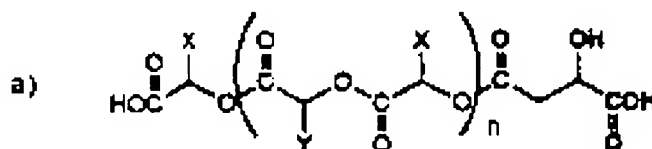


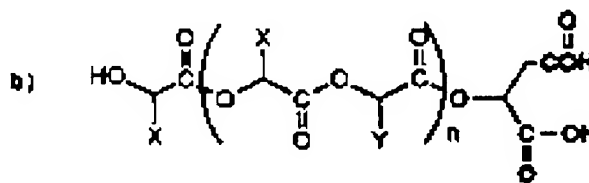
FIG. 3

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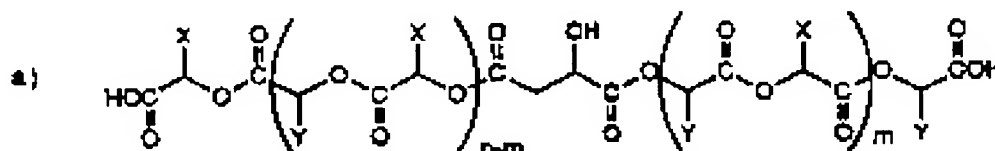
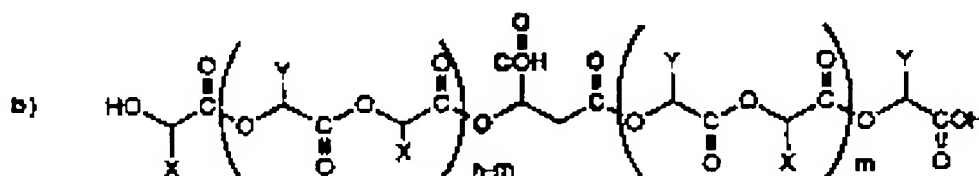
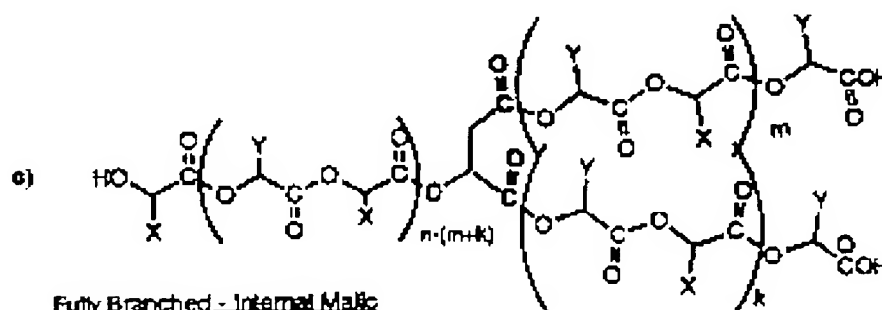
Forms of Lactate / Glycolate - Type Polymers of Polycarboxylic Acid-Tipped Chains

Di End Acid - Terminal Malic

I. Linear Chains

End Hydroxy - Terminal Malic

II. Branched Chains

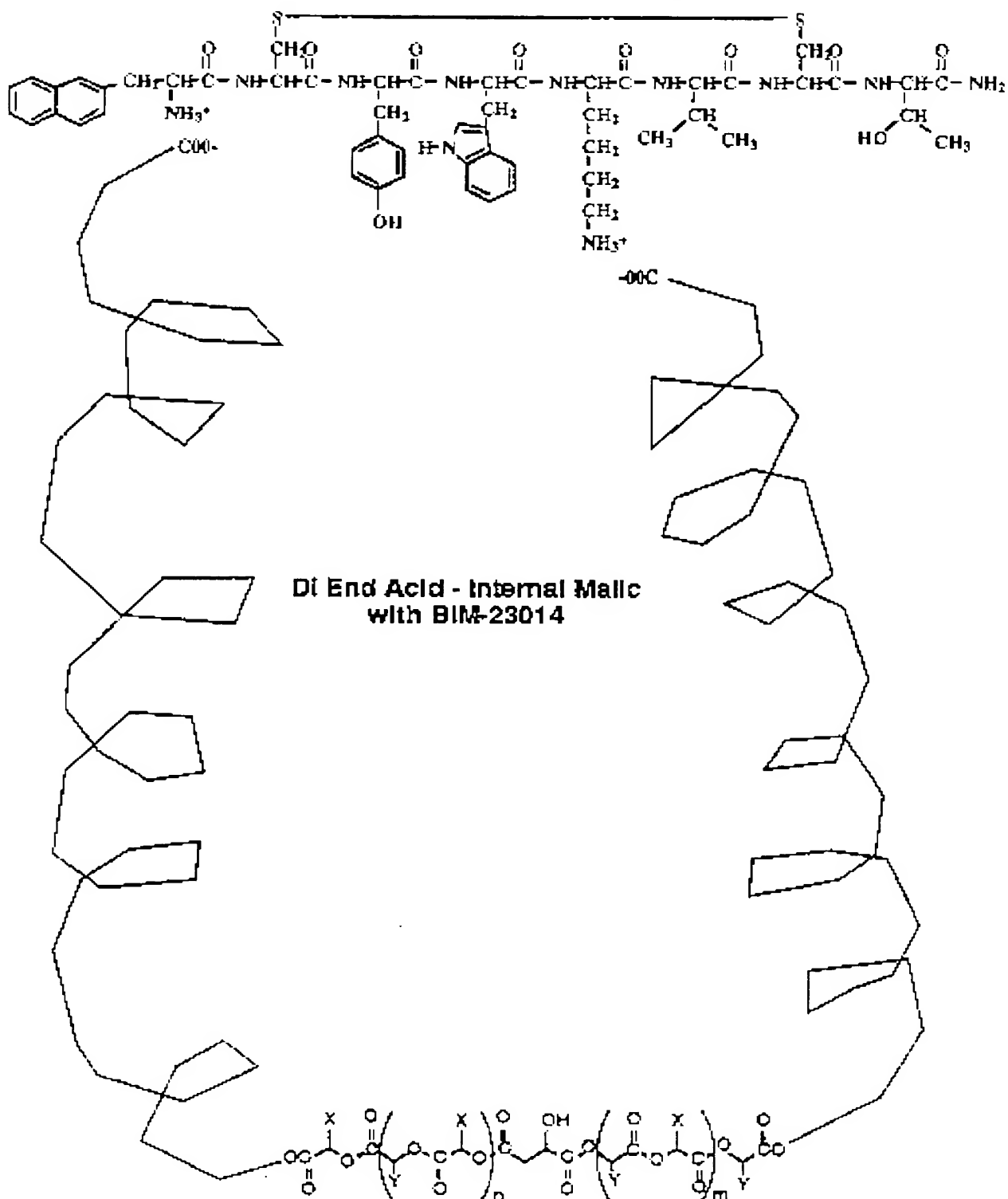
Di End Acid - Internal MalicEnd Hydroxy - Internal MalicFully Branched - Internal Malic

Y = CH₃ for Lactic, H for Glycolic; X = H for Glycolic, CH₃ for Lactic
Distribution of X + Y is formula dependent and polymerization dependent

FIG. 1

SUBSTITUTE SHEET (RULE 26)

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IONIC CONJUGATE EXAMPLE**FIG. 2****SUBSTITUTE SHEET (RULE 26)**

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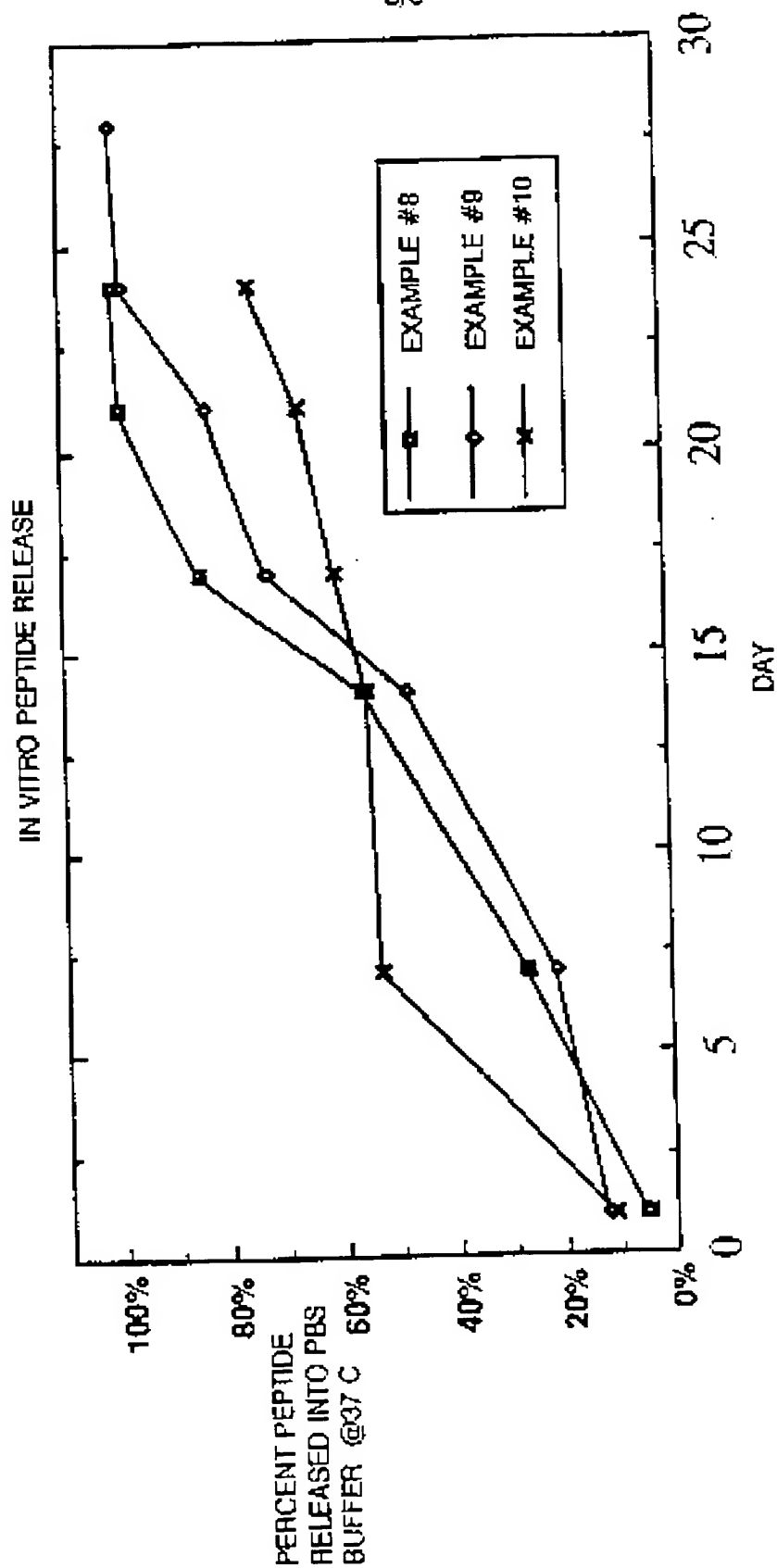


FIG. 3



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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(54) Title: IONIC MOLECULAR CONJUGATES OF BIODEGRADABLE POLYESTERS AND BIOACTIVE POLYPEPTIDES		
(57) Abstract Disclosed is a sustained release pharmaceutical composition. The composition includes a polyester containing a free COOH group ionically conjugated with a bioactive polypeptide comprising at least one effective ionogenic amine, wherein at least 50 % by weight, of the polypeptide present in the composition is ionically conjugated to the polyester.		

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/00148

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : A61K 9/16, 9/52, 37/02

US CL : 525/54.1; 530/815, 816

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 525/54.1; 530/815, 816

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS: (Polyester? or poly(w)ester? (p) (conjugate? or matrix or matrices) and (525*54.1/ccls or 530*815/ccls or 530*816/ccls)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X - Y	EP, A, 0 467 389 (UNIVERSITY OF KENTUCKY RESEARCH FOUNDATION) 22 January 1992. See page 4, lines 53, to page 5, line 23, for the polymer, page 5, lines 24-44, for the polypeptide. See Examples 1-7 for production of the conjugate.	1-3, 8-13 6, 7, 17, 21-30
X - Y	US, A, 5,162,505 (DEAN) 10 November 1992. See the Abstract, column 6, lines 11-44, and Example III.	1-3, 8-13 6, 7, 17, 21-30
X	US, A, 4,767,628 (HUTCHINSON) 30 August 1988. See column 2, line 55, to column 3, line 2, and Examples 1-35.	1-3, 8-13



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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"O" document referring to an oral disclosure, use, exhibition or other means		
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Date of the actual completion of the international search

26 MAY 1994

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

International application No.
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C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US, A, 4,356,166 (PETERSON) 26 October 1982. Note column 2, lines 51-64, for the carrier polymer and the Abstract.	4, 5, 14-16
Y	US, A, 4,609,707 (NOWINSKI) 02 September 1986. See column 8, lines 18-45 for the conjugates.	1, 2
Y	US, A, 5,071,909 (PAPPIN) 10 December 1991. See the Abstract, column 6, lines 17-35, and column 6, line 44 to column 8, line 44.	1-30

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